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## Dilution Buffer (Strengthen Blocking)

**Cat. No:** DB-02

**Pack Size :** 50mL/500mL

**For Research Use Only. Not For Use In Diagnostic Or Therapeutic Procedures**

### Applications

Serum and some samples often cause nonspecific binding, especially after blocking, which lead to high background and low sensitivity. The buffer can reduce non-specific adsorption in experiments, avoid background signals which is caused by serum especially, achieve high signal-to-noise ratio, and achieve ideal detection sensitivity.

It is for research use only.

### Shipping and Storage

Upon receipt, please store at 2-8°C, Once opened, store at 2-8°C.

### Example ELISA Procedure

#### 1. Materials and Reagents Preparation

Before starting the ELISA Assay, we should prepare the all reagents and materials required in the experiment. You can prepare these regents by following operations, **we provide the Dilution Buffer (Strengthen Blocking) (Cat. No. DB-02).**

Wash Buffer: PBS or TBS with 0.05% (v/v) Tween-20 (usually at pH7.4), 500 mL is sufficient for 96 tests. The pH of Buffer system can be adjust according to your experiment.

Substrate Dilution Buffer: 50 mM disodium hydrogen phosphate ( $\text{Na}_2\text{HPO}_4$ ) and 25 mM citric acid, adjust pH to 5.5 with 1 M Sodium hydroxide (NaOH), 25 mL is sufficient for 96 tests.

Substrate Stock Solution: 20 mg/mL TMB (Sigma-Aldrich, Catalog # 860336) in Dimethyl sulfoxide (Sigma-Aldrich, Catalog # D8418), 1 mL is sufficient for 96 tests. Protect from light TMB.

#### Substrate Working Solution

For each plate dilute 125  $\mu\text{L}$  substrate stock solution in 25 mL substrate dilution buffer and add 20  $\mu\text{L}$  5%  $\text{H}_2\text{O}_2$  (pipette 10  $\mu\text{L}$  30%  $\text{H}_2\text{O}_2$  into 50  $\mu\text{L}$  distilled water), mix well.

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**Notes:**

1) *The TMB Substrate Working Solution should be freshly prepared and used within 15 minutes.*

2) *If you choose to use other commercially available ready-to-use TMB substrate solutions, you should follow the manufacturer's instruction.*

Stop Solution: 1 M sulfuric acid (aqueous), 6 mL is sufficient for 96 tests.

Coated Plates (ACROBiosystems, Catalog # SP-11)

Microplate sealing film (Sigma-Aldrich, Catalog # Z724742)

Pipettes and pipette tips

UV/Vis microplate spectrophotometer (absorbance 450 nm, correction wavelength set to 630 nm)

## **2. Recommended Protocol**

### **2.1 Preparation**

Reconstitute and store all reagents as recommended.

### **2.2 Washing**

Add 300  $\mu$ L of Wash buffer to each well, gently tap the plate for 1 minute, remove any remaining Wash Buffer by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the wash step above for three times.

### **2.3 Add biotinylated protein or antibodies**

1) Dilute Biotinylated protein or antibodies to a concentration you want (usually 1~10  $\mu$ g/mL) with Dilution Buffer to make Biotinylated molecule working solution.

2) Add 100  $\mu$ L Biotinylated molecule to each well and incubate at 37°C or RT for 1 hour.

3) For Non - specific of the sample wells, please add 100  $\mu$ L Dilution Buffer.

### **2.4 Washing**

Remove the remaining solution by aspiration, add 300  $\mu$ L of Wash buffer to each well, gently tap the plate for 1 minute, remove any remaining Wash Buffer by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the wash step above for three times.

### **2.5 Add Samples**

1) Make series dilution of the samples as appropriate with Dilution Buffer.

2) Add 100  $\mu$ L of the serial dilution of sample to each well, incubate at 37°C or RT for 1 hour.

*If you have any questions, please contact our technical support team at: [TechSupport@acrobiosystems.com](mailto:TechSupport@acrobiosystems.com)  
<http://www.acrobiosystems.com>*

## 2.6 Washing

Repeat step 4.

## 2.7 Add primary antibody

- 1) Dilute primary antibody to an appropriate concentration with Dilution Buffer.
- 2) For all wells, add 100  $\mu$ L of diluted primary antibody, and incubate at 37°C or RT for 1 hour.

## 2.8 Washing

Repeat step 4.

## 2.9 Add enzyme-labeled secondary antibody

- 1) Dilute secondary antibody to an appropriate concentration with Dilution Buffer.
- 2) For all wells, add 100  $\mu$ L of diluted secondary antibody, and incubate at 37°C or RT for 1 hour, avoid light.

## 2.10 Washing

Repeat step 4.

## 2.11 TMB Substrate Reaction

Add 200  $\mu$ L TMB Substrate Working Solution to each well. Seal the plate with microplate sealing film and incubate at 37°C or RT for 20 minutes, avoid light.

## 2.12 Termination

Add 50  $\mu$ L Stop Solution to each well, and tap the plate gently for 3 minutes to allow thorough mixing.

*Note: the color in the wells should change from blue to yellow.*

## 2.13 Data Recording

Read the absorbance at 450 nm using UV/Vis microplate spectrophotometer.

*Note: the plate may be read at 600 nm without adding 1 M sulfuric acid, but the Signal-to-Background ratio.*

### 3. Example Data

#### Binding Assay between S protein RBD and Anti-SARS-CoV on SA Plate

Immobilized Biotinylated SARS-CoV-2 S protein RBD, His, Avi tag at 1000ng/mL (100  $\mu$ L/well) on Streptavidin Coated Plates, Clear, 96-Well (Cat. No. SP-11), can bind Anti-SARS-CoV antibody with a linear range of 0.1-3 ng/mL.

