

Influenza A (H1N1) Viruses Neuraminidase (NA) Specific ELISA Kit

Pack Size: 96 tests

Catalog Number: RAS-A223

IMPORTANT: Please carefully read this manual before performing your experiment.

For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedures



INTENDED USE

This kit is developed for specific quantitative detection of Influenza A (H1N1) viruses Neuraminidase (NA) in samples. It is intended for research use only (RUO).

PRINCIPLE OF THE ASSAY

Neuraminidase (NA) and hemagglutinin (HA) are major membrane glycoproteins found on the surface of influenza virus. Hemagglutinin binds to the sialic acid-containing receptors on the surface of host cells during initial infection and at the end of an infectious cycle. Neuraminidase, on the other hand, cleaves the HA-sialic acid bondage from the newly formed virions and the host cell receptors during budding. Neuraminidase thus is described as a receptor-destroying enzyme which facilitates virus release and efficient spread of the progeny virus from cell to cell. This assay kit is used to measure the levels of Influenza A (H1N1) viruses Neuraminidase (NA) protein by employing a standard sandwich-ELISA format. The microplate in the kit has been pre-coated with Anti-Influenza A (H1N1) Neuraminidase Antibody. First add the standard samples provided in the kit and your samples to the plate, incubate and wash the wells. Then add the HRP-Anti-Influenza A (H1N1) Neuraminidase Antibody to the plate, incubate and wash the wells. Lastly load the substrate into the wells and monitor color development in proportion with the amount of Influenza A (H1N1) viruses Neuraminidase (NA) protein present. The reaction is stopped by the addition of a stop solution and the intensity of the absorbance can be measured at 450 nm and 630 nm. The OD Value reflects the amount of Influenza A (H1N1) viruses Neuraminidase (NA) protein bound.

MATERIALS PROVIDED

TABLE 1. MATERIALS PROVIDED

Catalog	Catalog Components		Format	Storage	
J	•	tests)		Unopened	Opened
RAS223-C01	Pre-coated Anti-Influenza A (H1N1) Neuraminidase Antibody Microplate	1 plate	Solid	2-8°C	2-8°C
RAS223-C02	Influenza A [A/Victoria/4897/2022(H1N1)] Neuraminidase Standard	15 μg	Powder	2-8°C	-70°C
RAS223-C03	HRP-Anti-Influenza A (H1N1) Neuraminidase Antibody	20 μg	Powder	2-8°C, avoid light	-70°C, avoid light

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RAS223-C04	10×Washing Buffer	50 mL	Liquid	2-8°C	2-8°C
RAS223-C05	Dilution Buffer		Liquid	2-8°C	2-8°C
RAS223-C06	Substrate Solution	12 mL	Liquid	2-8°C, avoid light	2-8°C, avoid light
RAS223-C07	Stop Solution	7 mL	Liquid	2-8°C	2-8°C

REAGENTS/EQUIPMENT NEEDED BUT NOT SUPPLIED

Single or dual wavelength microplate reader with 450 nm and 630 nm filter;

Centrifuge;

37°C Incubator;

10 μL, 200 μL and 1000 μL precision pipettes;

 $10~\mu L$, $200~\mu L$ and $1000~\mu L$ pipette tips;

Multichannel pipettes;

Tubes;

Graduated cylinder to prepare Wash Solution;

Deionized or distilled water to dilute 10×Washing Buffer;

STORAGE

- 1. Unopened kit should be stored at 2°C-8°C upon receiving.
- 2. Find the expiration date on the outside packaging and do not use reagents past their expiration date.
- 3. The opened kit should be stored per components table. The shelf life is 30 days from the date of opening.

REAGENT PREPARATION

- 1. Bring all reagents and samples to room temperature (20°C-25°C) before use. If crystals have formed in buffer solution, place the sample in a 37 °C incubator until the crystals have completely dissolved and bring the solution back to room temperature before use.
- 2. Reconstitute the provided lyophilized materials to stock solutions with distilled, sterile water as recommended in Table 2 and place the materials for 15 to 30 minutes at room temperature with occasional gentle mixing. Avoid vigorous shaking. The reconstituted stock solutions should be stored at -70°C. It is recommended not to freeze-thaw

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more than 1 time, the packing specification shall not be less than 5 μg .

TABLE 2. RECONSTITUTION METHODS FOR 96 TESTS

ID	Components	Size	Stock Solution Con.	Reconstitution Buffer and Vol.
RAS223-C02	Influenza A [A/Victoria/4897/2022(H1N1)] Neuraminidase Standard	15 μg	150 μg/mL	100 μL water
RAS223-C03	HRP-Anti-Influenza A (H1N1) Neuraminidase Antibody	20 μg	200 μg/mL	100 μL water

RECOMMENDED SAMPLE PREPARATION

1. Working Fluid Preparation

1.1 Preparation of 1×Washing Buffer:

Dilute 50 mL 10×Washing Buffer with ultrapure water/deionized water to 500 mL.

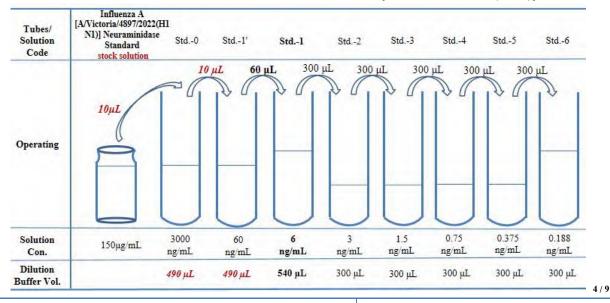
1.2 Preparation of HRP-Anti-Influenza A (H1N1) Neuraminidase Antibody working fluid:

Dilute HRP-Anti-Influenza A (H1N1) Neuraminidase Antibody to 0.2 μg/mL with Dilution Buffer. The prepared working fluid should avoid light. Please prepare it for one-time use only.

2. Preparation of Standard Curve

Make serial dilutions of the Influenza A [A/Victoria/4897/2022(H1N1)] Neuraminidase as a Standard curve with Dilution Buffer as recommended in Figure 1.

FIGURE 1. PREPARATION OF 1:1 SERIAL DILUTIONS OF THE Influenza A [A/Victoria/4897/2022(H1N1)] Neuraminidase



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ACTO*

3. Add Samples

Add 100µL serially diluted Influenza A [A/Victoria/4897/2022(H1N1)] Neuraminidase Standard curve and samples

to each well. For blank Control wells, please add 100µL Dilution Buffer. Seal the plate with microplate sealing film and

incubate at room temperature for 1.0 hour.

Note: It is recommended to set double holes for samples and standard curves to be tested.

4. Washing

Remove the remaining solution by aspiration, add 300 µL of 1×Washing Buffer to each well, soak for 30s, remove any

remaining 1×Washing Buffer: by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the

wash step above for three times.

5. Add HRP-Anti-Influenza A (H1N1) Neuraminidase Antibody

For all wells, add 100 µL HRP-Anti-Influenza A (H1N1) Neuraminidase Antibody (dilute to 0.2 µg/mL) working

solution. Seal the plate with microplate sealing film and incubate at room temperature for 1.0 hour.

6. Washing

Repeat step 4.

7. Substrate Reaction

Add 100 µL Substrate Solution to each well. Seal the plate with microplate sealing film and incubate at room

temperature for 20 min, avoid light.

8. Termination

Add 50 µL Stop Solution to each well and tap the plate gently to allow thorough mixing.

Note: the color in the wells should change from blue to yellow.

9. Data Recording

Read the absorbance at 450 nm and 630 nm using UV/Vis microplate spectrophotometer within 5 minutes.

Note: To reduce the background noise, subtract the value read at $OD_{450 \text{ nm}}$ with the value read at $OD_{630 \text{ nm}}$.

CALCULATION OF RESULTS

1. Normal range of Standard curve: R²≥0.9900, detection range: 0.188-6 ng/mL.

2. If the OD value of the sample to be tested is higher than the highest standard, the sample shall be diluted with

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dilution buffer and assay repeated.

3. To calibrate absorbance value obtained by the standard curve, the OD value of the sample to be measured is subtracted from the OD value of the blank control. The standard curve is plotted with the standard concentration as x-axis and the calibrated absorbance value as y-axis. Four parameters logistic are used to draw the standard curve and calculate the sample concentration.

PRECAUTIONS

- 1. This kit is for research use only and is not for use in diagnostic or therapeutic procedures.
- 2. The kit should be used according to the instructions.
- 3. Do not mix reagents from different lots.
- 4. Bring all reagents and samples to room temperature (20°C-25°C) before use. If crystals have formed in buffer solution, warm to room temperature until the crystals have completely dissolved.
- 5. The kit should be stored at 2°C to 8°C.

TYPICAL DATA

The following data is for reference only. The sample concentration was calculated based on the results of the standard curve.

Standard (ng/mL)	O.D1	O.D2	Average	Corrected	25
6	2.457	2.503	2.480	2.471	R ² =0.9999
3	1.392	1.495	1.444	1.434	
1.5	0.725	0.761	0.743	0.734	0 Dptical Density
0.75	0.396	0.390	0.393	0.384	8 1
0.375	0.202	0.207	0.205	0.195	0.5
0.188	0.110	0.111	0.111	0.101	<i>y</i>
0	0.009	0.010	0.010	1	Conc.(ng/mL)



PRECISION

1. Intra-assay Precision

Three samples of known concentration were tested ten times on one plate to assess intra-assay precision.

2. Inter-assay Precision

Three samples of known concentration were tested in three separate assays to assess inter-assay precision.

	Intra-assay Precision]	Inter-assay Precision	n
Sample	1	2	3	1	2	3
n	10	10	10	3	3	3
Mean (ng/mL)	4.363	1.006	0.500	4.325	0.978	0.489
SD	0.108	0.015	0.017	0.043	0.031	0.027
CV (%)	2.5	1.5	3.3	1.0	3.2	5.6

Note: The example data is for reference only.

RECOVERY

Three samples with different concentrations were tested to calculate the recovery rate.

Sample(n=5)	Average Recovery %	Range %
High	99.5	88.0-106.1
Middle	100.7	92.1-109.8
Low	105.5	96.1-115.8

LINEARITY

To assess the linearity of the assay, samples spiked with high concentrations were serially diluted with calibrator diluent to produce samples with values within the dynamic range of the assay.

		Cell culture medium (DMEM)	Cell culture medium (1640)	
1.2	Average Recovery (%)	93.5	91.8	
1:2 Range (%)		91.4-96.3	88.5-99.3	
1:4	Average Recovery (%)	95.7	93.3	

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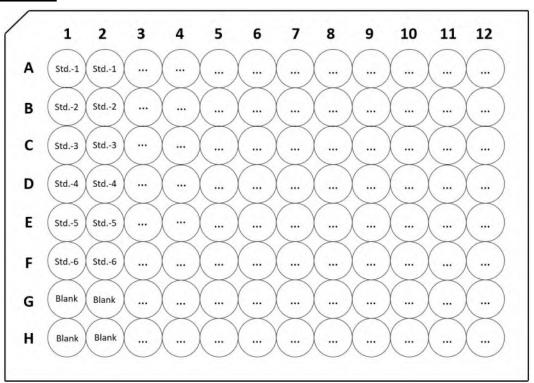


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	Range (%)	91.8-97.3	92.0-96.6
1:8	Average Recovery (%)	103.1	97.0
1:8	Range (%)	100.8-105.5	94.5-101.5
1.16	Average Recovery (%)	109.8	99.0
1:16	Range (%)	108.3-111.3	92.5-104.3

Note: The example data is for reference only.

PLATE LAYOUT



Note: Blank is a Blank Dilution Buffer hole.





TROUBLESHOOTING GUIDE

Problem	Cause	Solution	
Poor standard curve	* Inaccurate pipetting	* Check pipettes	
Lauga CV	* Inaccurate pipetting	* Check pipettes	
Large CV	* Air bubbles in wells	* Remove bubbles in wells	
High hooligwoond	* Plate is insufficiently washed	* Review the manual for proper wash.	
High background	* Contaminated wash buffer	* Make fresh wash buffer	
Very low readings across the	* Incorrect wavelengths	* Check filters/reader	
plate	* Insufficient development time	* Increase development time	
Samples are reading too high, but standard curve looks fine	* Samples contain cytokine levels above assay range	* Dilute samples and run again	
		* Assay set-up should be continuous - have all standards	
		and samples prepared appropriately before commencement	
Drift	* Interrupted assay set-up	of theassay	
Din	* Reagents not at room temperature	* Ensure that all reagents are at room temperature before	
		pipetting into the wells unless otherwise instructed in the	
		antibody inserts	

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